Development and evaluation of a novel product to remove surface contamination of hazardous drugs



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Abstract

Background: Even while following best practices, surface exposures of hazardous drugs (HDs) are high and numerous. Thus, it is important to develop new products to reduce the surface contamination of HDs. Hazardous Drug Clean (HDCleanTM) was developed to decontaminate and remove HDs from various types of surfaces and overcome the problems associated with other cleaning products.

Methods: HDClean was evaluated to remove mock surface exposures of HDs (docetaxel, paclitaxel, ifosfamide, cyclophosphamide, 5-FU, and cisplatin) from various types of surfaces. In two separate cancer centers, studies were performed to evaluate HDClean in reducing surface contamination of HDs in the pharmacy departments where no closed system transfer device (CSTD) was used. In a third cancer center, studies were performed comparing the effectiveness of a CSTD + Surface Safe compared with CSTD + HDClean to remove HDs.

Results: HDClean was able to completely remove mock exposures of a wide range of HDs from various surfaces (4 and 8 sq ft areas). Daily use of HDClean was equal to or more effective in reducing surface contamination of HDs in two pharmacies compared with a CSTD. HDClean was significantly more effective in removing HDs, especially cisplatin, compared with Surface Safe and does not have the problems associated with decontamination solutions that contain sodium hypochlorite.

Conclusion: These studies support HDClean as an effective decontaminating product, that HDClean is more effective than Surface Safe in removing HDs and is equal to or more effective than CSTD in controlling HD surface exposures.

Keywords

Hazardous drugs, surface contamination, decontamination, cleaning, HDClean

Introduction

Medications play an important role in treating and managing various disease states. With their broad use, numerous providers, patients, and family members could have contact with these drugs. For disease states such as organ transplantation, autoimmune diseases, cancer, and infections, the specific medications used for management have been demonstrated to exhibit specific concerns for the health care worker.¹ Studies over the past five decades show employees handling hazardous drugs (HDs) are at increased risk for drug-related toxicities.^{2–11} HDs may exert undesirable effects on employees through direct skin contact or systemic

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absorption due to the inhalation of aerosols, hand-tomouth transfer, or accidental needle sticks.

Even with appropriate aseptic technique, work area surfaces may easily become contaminated with HDs leading to undesirable occupational exposure.¹⁰ One study published in 1979 described the occupational adverse effects of handling HDs. Researchers found significantly increased mutagenicity of urine bacteria concentrates in nurses handling antineoplastic agents when compared to controls.¹² In a more recent crosssectional study of 68 exposed health care workers and 53 controls at three US-based cancer centers, measurable concentrations of antineoplastic agents were detected in urine samples from health care workers after chemotherapy preparation.¹³

At the base of occupational exposure prevention methods for HDs are engineering controls, proper procedures, and personal protective equipment (PPE). Besides proper aseptic technique training, one must also utilize workplace controls to minimize exposure to HDs. Primary engineering controls, a Compounding Aseptic Containment Isolator (CACI) or a Class II Biological Safety Cabinet (BSC) Type B2, should be used for all compounding activities, being physically separated from non-hazardous compounding areas.¹⁴ PPE is intended to minimize exposure to health care workers by providing a barrier between a worker and the hazardous compound. These include appropriately rated and chemotherapy-tested gloves, gowns, and facemasks when necessary. In addition, many organizations have implemented closed system transfer devices (CSTDs). Since the first device became available in 1997 (PhaSeal[®]), there has been a steady rise in utilization within the health care setting. CSTDs are drug-transferring devices mechanically prohibiting the transfer of environmental contaminants into the sterile system and the escape of hazardous drugs or vapors outside the system.¹⁵ Unites States Pharmacopeia (USP) recommends the use of CSTDs, primarily to decrease the exposure to HDs.¹⁶ However, studies suggest that an additional source of HD surface contamination is attributed to exposures of HDs on the outsides of vials generated during the manufacturing process.^{17,18}

Healthcare workers who are exposed to HD as part of their work practice should take precaution to eliminate or reduce exposure as much as possible. Nurses and pharmacists who prepare and/or administer these hazardous drugs are the two occupational groups who have the highest exposure potential to hazardous drugs. Studies designed to detect chemotherapy surface contamination show that 80–90% of nursing and pharmacy related sites evaluated for HD surface residue have at least one area of detectable hazardous drug surface contamination, even while following best practices.^{19–28} Because of this, it is important to evaluate and develop further methods to reduce and remove the surface contamination of HD.

The draft of USP800 (May 2015) includes recommendations and best practices on ways to prevent and to remove surface contamination of HDs.²⁹ Potential methods of removal of HDs include deactivation, decontamination and cleaning. Deactivation renders a compound inert or inactive. Decontamination occurs by physically removing HD residue from non-disposable surfaces and transferring it to absorbent, disposable materials (e.g. wipes, pads, or towels) appropriate to the area being cleaned. All disposable materials must be discarded as contaminated HD waste. As related to deactivation, chemical deactivation of HD residue is preferred, but no single process has been found to deactivate all currently available HDs. Studies have examined oxidizing agents that have varying results. Moreover, some potential deactivators have produced byproducts that are as hazardous as the original drug. Other deactivators have respiratory effects or result in caustic damage to surfaces. For example, sodium hypochlorite is corrosive to stainless steel surfaces if left untreated; therefore, sodium hypochlorite must be neutralized with sodium thiosulfate or followed by use of a germicidal detergent. Cleaning is a process that results in the removal of contaminants (e.g. HD residue) from objects and surfaces using water, detergents, surfactants, solvents, and/or other chemicals. A multicomponent system to remove surface contamination of HDs is theoretically more efficient than a single component or method system because of the diverse nature of HDs. For example, a product that combines the ability to decontaminate and clean may be most effective. With the availability of assays to measure HD surface contamination, USP800 recommends surface wipe sampling to document the effectiveness of any agent used remove HD residue from work surfaces.

Surface Safe is a towelette system containing sodium hypochlorite that is designed to decontaminate and deactivate surface contamination of some HDs.³⁰ However, studies evaluating Surface Safe did not evaluate or report the removal of HDs or evaluate exposures of HDs on surfaces. Moreover, the studies were only performed in test tubes and only evaluated if the form of the 3 HDs with very similar chemical characteristics was potentially changed by evaluating binding to DNA. Thus, formal studies evaluating the ability of this product to remove a wide range of HDs from surfaces have not been reported. In addition, Surface Safe is associated with a very strong odor due to the sodium hypochlorite, leaves an oil-like residue that must be removed with the use of an additional cleaning product, and is designed to work on a relatively small area (2 sq ft area). Moreover, the sodium hypochlorite in Surface Safe is corrosive to stainless-steel surfaces.

Hazardous Drug Clean (HDCleanTM) was developed to address the need for a product that can decontaminate, clean and remove HDs with highly variable chemical characteristics from various types of surfaces. In addition, HDClean is a two-step towelette cleaning product that was designed to remove HDs from surfaces and to overcome the problems associated with Surface Safe (e.g. odor, corrosiveness, need for second cleaning product, use of a small area). Here, we report a series of research and development studies and testing in three separate cancer centers that evaluated the ability of HDClean to remove surface contamination of a series of HD with vastly different chemical characteristics and solubilities. The evaluation of HDClean was performed in pharmacies and nursing units. Analytical chemistry methods also were used to accurately measure the concentrations of HDs on surfaces as outlined in the draft of USP800 (May 2015).²⁹

Methods

Evaluation of HDClean on mock surface contaminations

Objectives. The goal of this experimentation was to evaluate the effectiveness of HDClean in removing surface contamination of HDs from a stainless steel surface. HDClean was used to clean 4 sq ft ($2 \text{ ft} \times 2 \text{ ft}$) and 8 sq ft ($4 \text{ ft} \times 2 \text{ ft}$) areas. The HDClean dual component towelettes were used to clean all areas using the standard cleaning procedure. HDClean towelette #1 (quaternary ammonium based solution) was used to clean each area first and then HDClean towelette #2 (isopropyl alcohol-based solution) was used to clean each area. This procedure was repeated a second time on each contaminated area.

Study design. Docetaxel, paclitaxel, ifosfamide, cyclophosphamide, and 5-fluorouracil (5-FU) were added to a single solution in 37.5:37.5:25 methanol:acetonitrile:water to give a final nominal concentration of 500 ng/mL of each drug. Cisplatin was added to a separate solution of 2% nitric acid to give a 500 ng/mL solution. Surfaces were "contaminated" with the drug solutions of 500 ng/mL to give a nominal amount of 500 ng of each drug per square foot. All solutions were allowed to dry completely before testing. These drugs were selected as they represent some of the most frequently administered anticancer agents and have a wide variety of chemical characteristics (e.g. structures, solubilities).

Cleaning and sampling procedures. HDClean was used to clean 4 and 8 sq ft areas using the standard methods described above. After cleaning, each area was then sampled (n=4 separate $1 \text{ ft} \times 1 \text{ ft}$ areas) using ChemoGLO (Chapel Hill, NC) wipe kits to extract all six drugs from the surface. Surface areas that were not treated with HDClean were also sampled $(n=4 \text{ separ$ $ate } 1 \text{ ft} \times 1 \text{ ft}$ areas) as baseline measurements of surface exposures of HD. ChemoGLO wipe kits and analytical methods were used to evaluate the surface exposure of docetaxel, paclitaxel, 5-FU, ifosfamide, cyclophosphamide and cisplatin as described below in detail.

Evaluation of HDClean on different types of surfaces

Objectives and study design. This investigation was designed to evaluate the effectiveness of HDClean at removing HD contamination on different types of surfaces over the course of one week. Six different areas were selected based on frequency of use and potential for contamination or exposure in pharmacies and nursing units. A summary of the surface areas and materials that were evaluated is given in Table 1. These studies were performed in a separate hospital than described below for Cancer Centers #1, #2, and #3.

Study design, cleaning and sampling procedures. A section of each area measuring 4 sq ft $(2 \text{ ft} \times 2 \text{ ft})$ was delineated. This large section was then subdivided into four 1 sq ft sections. Before cleaning, initial drug contamination was evaluated using ChemoGLO wipe kits to sample 1 square foot of each area. HDClean wipes were used as directed to clean the six selected surfaces daily at end of the day. Surface contamination of HD was evaluated at end of the second and fifth day of cleaning with HDClean. ChemoGLO wipe kits were used to evaluate surface exposure of docetaxel, the paclitaxel. 5-FU, ifosfamide, cyclophosphamide and platinum analogues (e.g. cisplatin, carboplatin, and oxaliplatin) as outlined below.

Evaluation of HDClean in pharmacy at cancer centers #1 and #2 not using a CSTD

Objective. The primary objective of this study was to evaluate the effectiveness of HDClean in reducing

Table	١.	Summary	of	surface	area	and	material	evaluated.

Surface area	Material
BSC	Stainless steel
Floor under BSC	Waxed tile
Table top	Phenolic resin
Floor under preparation area	Waxed tile
Computer keyboard	Plastic; phenolic resin

surface contamination of HDs in the pharmacy departments of two separate cancer centers where no CSTD or robot was used to prepare doses of chemotherapy.

Study design, cleaning, and sampling procedures. Chemotherapy surface contamination was evaluated in both cancer centers before and after the use of HDClean. Six different locations in each cancer center were evaluated for surface contamination of docetaxel, paclitaxel, 5-FU. cvclophosphamide, ifosfamide, methotrexate and platinum analogues. Surface samples were obtained at baseline (prior to cleaning with HDClean). HDClean was used twice per day (in the morning and afternoon) in all studies. In study 1, wipe studies were performed on day 5 of weeks 1 and 2 after using HDClean in the morning, preparing chemotherapy during the day but before performing the second HDClean cleaning procedure in the afternoon (first HDClean cleaning procedure \rightarrow preparing chemotherapy \rightarrow surface wipe study \rightarrow second HDClean cleaning procedure). In study 2, the wipe studies were performed on day 5 of week 1 after preparation of chemotherapy and immediately following the second daily cleaning procedure using HDClean (first HDClean cleaning procedure \rightarrow preparing chemotherapy \rightarrow second HDClean cleaning procedure \rightarrow surface wipe study). Surface contamination of HD was evaluated using ChemoGLO wipe kits. The wipe Kit samples were analyzed for docetaxel, paclitaxel. 5-FU, cyclophosphamide, ifosfamide, platinum analogues methotrexate and at the ChemoGLO reference lab as described below.

Evaluation of HDClean in pharmacy and nursing unit at cancer center #3

Objectives. The studies outlined above evaluated HDClean in pharmacy departments that prepare chemotherapy. However, there is also the potential for high exposures of HDs in nursing units where chemotherapy is administered to patients. Thus, this study evaluated the effectiveness of HDClean to reduce exposures of HD on surfaces in a pharmacy and a nursing unit in a separate cancer center from the studies performed in cancer centers #1 and #2. In addition, the effectiveness of HDClean in removing surface contamination of HDs was compared to Surface Safe.

Study design, cleaning and sampling procedures. Studies were performed to evaluate the ability of HDClean to remove chemotherapeutic agents from surfaces in the pharmacy department and nursing unit in a cancer center. The center utilized all best practices, including a CSTD and Surface Safe[®], a commercially available decontamination agent. Surface Safe[®] was used as per

manufacturer's recommendations. Six surface wipe studies were conducted using the ChemoGLO wipe kit, four in pharmacy and two in nursing unit. Surface wipe studies were performed at baseline when Surface Safe + CSTD were used and after changing the cleaning product to HDClean (HDClean + CSTD). HDClean was used daily for two weeks. The ChemoGLO wipe Kit samples were analyzed for docetaxel, paclitaxel, 5-FU, cyclophosphamide, ifosfamide, and methotrexate at the ChemoGLO reference lab as described below.

Surface sampling wipe kits and analytical assay

ChemoGLO Wipe KitsTM (Chapel Hill, NC) quantifies amounts of contaminants of the antineoplastic agents 5-FU, ifosfamide, cyclophosphamide, methotrexate, docetaxel, paclitaxel and platinum analogues (e.g. cisplatin, carboplatin and oxaliplatin) on surfaces. Each kit contains enough materials to conduct six wipes. To ensure proper technique in collecting the samples, a training video and written instructions were provided by the reference lab and in turn reviewed by the research team that collected the samples. Standardized surfaces of approximately $144 \, {\rm in}^2$ $(12 \times 12 \text{ inches})$ were sampled with two wipes each, one vertical wipe and one horizontal wipe. Prior to wiping, each swab was moistened with 2 mL of a solution containing isopropyl alcohol. The wipe procedure is able to recover >90% of cyclophosphamide, ifosfamide, paclitaxel, docetaxel, 5-FU and methotrexate from a 12×12 inch surface.

Samples were stored at 4°C on site until they were shipped to the ChemoGLO reference lab where they were stored at 4°C until processed and analyzed. A 200 µL solution containing internal standards (IS) [cyclophosphamide-d4 (CPM-d4) for cyclophosphamide; antipyrine (APR) for ifosfamide; paclitaxel-d5 paclitaxel; docetaxel-d9 for docetaxel; for 5-FU-¹³C,-¹⁵N₂ for 5-FU; methotrexate-d3 for methotrexate] was added to each swab as the internal standard. The drugs and IS were extracted from the wipe swab using 2mL of an extraction solution. The contents (swab and solution) were transferred to a Salivette tube with an insert and centrifuged at 4000 rpm for 10 min. A 200 µL aliquot of the resulting solution was removed from the bottom chamber of the Salivette tube, dried down and then reconstituted with $30\,\mu\text{L}$ of mobile phase solution. The sample was then analyzed by liquid chromatography-tandem mass spectrometry (LC-MS/ MS) assays using an Agilent 6410 Triple Quadrupole as previously described.^{31–35} The range of cyclophosphamide, ifosfamide, paclitaxel, docetaxel, 5-FU and methotrexate concentrations measured by the assays was linear 10 to 2000 ng/mL per swab area. The

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Experiment		Docetaxel	Paclitaxel	5-FU	lfosfamide	Cyclophos- phamide	Cisplatin
Baseline no cleaning [I ft \times I ft = I sq ft area] $(n = 7)^{a}$	Drug Amt (ng/sq ft)	525.5 ± 150.0	562.6±68.2	596.8±25.5	521.2±75.1	486.4±31.1	479.5±80.3
	No. of detectable samples	n = 4	n = 4	n = 4	n = 4	n = 4	n = 4
HDClean [2 ft \times 2 ft = 4 sq ft area] $(n = 10)^{a}$	Drug amount (ng/sq ft)	ND	ND	ND	ND	ND	ND
	No. of detectable samples	n = 0	n = 0	n = 0	n = 0	n = 0	n = 0
HDClean double area [2 ft \times 4 ft = 8 sq ft area] $(n = 12)^{a}$	Drug amount (ng/sq ft)	ND	ND	7.3 ± 13.8	17.6±31.9	ND	187.5±73.2
	No. of detectable samples	<i>n</i> = 0	n = 0	n = 4	n = 3	n = 0	n = 12

Table 2. Mean \pm SD concentrations (ng/sq ft) of six antineoplastic drugs on surfaces before and after cleaning procedures in the mock surface contamination studies.

ND = non-detectable drug exposure.

^aThe number listed represents the number of replicate areas sampled for surface contamination at base (n = 7), HDClean 4 sq ft area (n = 10) and HDClean 8 sq ft area (n = 12). The number of replicate sampling areas was increased from baseline to HDClean 4 sq ft area to HDClean 8 sq ft area in order to get a better representation of the exposures over the entire areas.

correlation coefficients for three successive duplicate standard curves were all >0.99. The accuracy for each drug at the lower limit of quantitation (LLQ) and for the standard concentrations were <20% and <15%, respectively, of the theoretical values. Intra-assay and inter-assay precision was <10% for all drugs. As the volume of solutions added to each swab is maintained constant, the results are reported as ng/sq ft. The results of the two swabs used to wipe each area (1 swab used to wipe horizontal and 1 swab used to wipe vertical) are then added together to give a final exposure result for each sample area in ng/sq ft.

The same swab used for the surface sampling of the drugs listed above underwent further sample processing to measure platinum (Pt) analogues via Inductively Coupled Plasma Mass Spectrometry (ICP-MS).^{36,37} The swab was processed as previously described.^{36,37} Briefly, the swab in processed with 1 mL of 5% of nitric acid that contains 20 ppb of Iridium (Ir) as the internal standard (IS), followed by 1 mL of water. All washes from each swab are pooled to produce 2 mL of reconstituted sample in 2.5% nitric acid in water. Analysis of the samples is performed using a Nexion 300 D ICP-MS, in which elemental metals are ionized through heating and then drawn into a mass spectrometer for detection based on mass-to-charge ratios (m/z). Pt concentrations measured by the assays were linear

10–1,000 ng/mL per swab area. The accuracy for each drug at the lower limit of quantitation (LLQ) and for the standard concentrations was <20% and <15%, respectively, of the theoretical values. Intra-assay and inter-assay precision were <10% for all drugs.

Results

Evaluation of HDClean on mock surface contaminations

HDClean was evaluated to remove the surface contamination of docetaxel, paclitaxel, 5-FU, ifosfamide, cyclophosphamide, and cisplatin that was added to the surfaces to achieve a final exposure of 500 ng/sq ft on 4 sq ft and 8 sq ft areas. The results of this study are summarized in Table 2. The baseline (prior to cleaning) surface exposure of each drug was approximately 500 ng/sq ft. The use of HDClean on the 4 sq ft area resulted in non-detectable concentrations of all drugs in all 10 surface areas. The use of HDClean on the 8 sq ft area resulted in non-detectable concentrations of docetaxel, paclitaxel and cyclophosphamide in all 12 surface areas. HDClean also achieved pronounced reductions in 5-FU and ifosfamide exposures on the 8 sq ft area. The mean % reduction in 5-FU was 98.8% with detectable drug in only 4 of the 12 surface areas.

Surface area cleaned		Docetaxel exposure at baseline (prior to cleaning)	Docetaxel exposures after 2 daily cleanings with HDClean	Docetaxel exposures after 5 daily cleanings with HDClean
BSC	ng/sq ft (% reduction)	34.7	ND (100%)	ND (100%)
Floor under BSC	ng/sq ft (% reduction)	80.5	37.6 (53.3%)	ND (100.%)
Table top	ng/sq ft (% reduction)	>4000.0	3848.6 (3.8%)	411.9 (89.7%)
Floor under preparation area	ng/sq ft (% reduction)	310.5	88.6 (71.5%)	58.0 (81.3%)
Computer keyboard	ng/sq ft (% reduction)	4.	ND (100%)	ND (100%)

Table 3. S	Summary of	Docetaxel	surface ex	kposures at	: baseline	and after	HDClean.
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ND = non-detectable drug exposure.

Table 4. Summary of paclitaxel surface exposures at baseline and after HDClean.

Surface area cleaned		Paclitaxel exposure at baseline (prior to cleaning)	Paclitaxel exposures after two daily cleanings	Paclitaxel exposures after five daily cleanings
BSC	ng/sq ft (% reduction)	19.9	ND (100%)	ND (100%)
Floor under BSC	ng/sq ft (% reduction)	ND	ND	ND
Table Top	ng/sq ft (% reduction)	>4000.0	311.4 (92.2%)	73.3 (98.2%)
Floor under preparation area	ng/sq ft (% reduction)	372.9	31.4 (91.6%)	30.2 (81.3%)
Computer keyboard	ng/sq ft (% reduction)	22.9	ND (100%)	ND (100%)

ND = non-detectable drug exposure.

The mean % reduction in ifosfamide was 96.6% with detectable drug in only 3 of 12 surface areas. In addition, HDClean achieved pronounced reductions in cisplatin in all 12 sample areas evaluated with mean percentage reduction of 60.9%. In summary, HDClean was equally effective in removing all drugs from the 4 sq ft and 8 sq ft areas. The only exception was for cisplatin where HDClean resulted in a 100% and 60% reductions in the 4 sq ft and 8 sq ft areas, respectively.

Evaluation of HDClean on different types of surfaces

The ability of HDClean to remove HDs from different types of surfaces was evaluated after 2 and 5 daily cleanings. The types of surfaces evaluated are summarized in Table 1 and included BSC, floors, bench tops and computer key boards. The summary of surface exposures docetaxel, paclitaxel, 5-FU and cisplatin at baseline and after 2 and 5 daily cleanings are included in Tables 3–6. The baseline exposures of the drugs were

highly variable on the different types of surfaces. Daily of use HDClean after two days and especially five days resulted in pronounced reductions of all drugs on all types of surfaces with most surfaces have nondetectable exposures of HD after use of HDClean. Ifosfamide was only detectable at baseline on table top (141.3 ng/sq ft) and floor under preparation (19.6 ng/sq ft) area. Both locations were non-detectable after two daily cleanings using HDClean. Cyclophosphamide was only detectable at baseline in the biological safety cabinet (BSC) (620.1 ng/sq ft). After two and five daily cleanings with HDClean, the exposures of cyclophosphamide were 197.6 ng/sq ft (68.2% reduction) and non-detectable (ND) (100.0% reduction). In most cases, all of the drugs were non-detectable on all surfaces after use of HDClean for five consecutive days.

Evaluation of HDClean in pharmacy at cancer center #1 and #2 not using a CSTD

This study evaluated the surface exposure of chemotherapy in two separate cancer centers that were not

Surface area cleaned		5-FU exposure at baseline (prior to cleaning)	5-FU exposures after two daily cleanings	5-FU exposures after five daily cleanings
BSC	ng/sq ft (% reduction)	172.9	47.9 (72.1%)	ND (100%)
Floor under BSC	ng/sq ft (% reduction)	2285.0	266.2 (88.4%)	247.8 (89.16%)
Table Top	ng/sq ft (% reduction)	>4000.0	475.5 (88.1%)	163.1 (95.9%)
Floor under preparation area	ng/sq ft (% reduction)	>4000.0	226.4(94.3%)	186.3 (95.3%)
Computer keyboard	ng/sq ft (% reduction)	384.6	ND (100%)	ND (100%)

Table 5. Summary of 5-FU surface exposures at baseline and after HDClean.

ND = non-detectable drug exposure.

Table 6. Summary of cisplatin surface exposures at baseline and after HDClean.

Surface area cleaned		Cisplatin exposure at baseline (prior to cleaning)	Cisplatin exposures after two daily cleanings	Cisplatin exposures after five daily cleanings
BSC	ng/sq ft (% reduction)	20.1	ND (100%)	ND (100%)
Floor under BSC	ng/sq ft (% reduction)	22.9	ND (100%)	ND (100%)
Table Top	ng/sq ft (% reduction)	423.9	16.6 (96.1%)	ND (100%)
Floor under preparation area	ng/sq ft (% reduction)	34.9	ND (100%)	ND (100%)
Computer keyboard	ng/sq ft (% reduction)	24.4	ND (100%)	ND (100%)

ND = non-detectable drug exposure.

using a CSTD. In study 1, wipe studies were performed on day 5 of weeks 1 and 2 after using HDClean in the morning, preparing chemotherapy during the day but before performing the second HDClean cleaning procedure in the afternoon on that day. In study 2, the wipe studies were performed after preparation of chemotherapy and immediately following the second daily cleaning procedure using HDClean.

A summary of surface exposures of the HDs at baseline and after use of HDClean in studies 1 and 2 our presented in Tables 7 (cancer center #1) and 8 (cancer center #2). In study 1, the concentrations of most drugs at most locations at 1 and 2 weeks after starting HDClean achieved pronounced reductions in exposures as compared to baseline even though the wipe studies were performed after the preparation of chemotherapy on that day but before the second HDClean procedure.

In study 2, the surface exposure of all drugs on all surfaces were non-detectable except for 5-FU in one location in cancer center #2. Thus, the total percentage of detectable exposures of drugs in study 2 was only 1.1% [1 out of 84 (7 drugs in 12 locations) potential drug exposures]. Based on the results of study 2, the few high or increased concentrations of drugs in study 1 on weeks 1 and 2 compared to baseline are most likely due to surface contamination associated with preparing chemotherapy that day as the wipe studies were performed prior use of HDClean. These results highlight the potential of HD accumulation on a daily basis.

Evaluation of HDClean in pharmacy and nursing unit at cancer center #3

Surface exposures of chemotherapy were measured in both pharmacy and nursing units for five drugs in a total of six locations for a total of 30 sample results. These studies were performed in a separate cancer center from sites #1 and #2. The baseline wipes were performed after use of CSTD and Surface Safe. The cancer center then switched to the use of HDClean to clean surfaces and wipe studies. Wipe studies after the implementation of HDClean were then performed in the same locations as the baseline studies.

Surface exposures of chemotherapeutic agents prior to and after the use of HDClean in the pharmacy and nursing units are presented in Table 9. In wipe studies prior to the use of HDClean, there were detectable exposures for four of the five drugs analyzed with drugs detectable in 8 of 30 samples (26.7%). In addition, paclitaxel and ifosfamide exposures were >600 ng/sq ft at baseline. The baseline results are especially important as the site was using a CSTD and the

Table 7. Surfac	ce exposure c	Table 7. Surface exposure of chemotherapeutic agents at baseline	and in studies	I and 2 after	use of HDCle	baseline and in studies 1 and 2 after use of HDClean in cancer center $\#1$.			
			Docetaxel exposure	Paclitaxel exposure	5-FU exposure	Cyclophos- phamide	lfosfamide exposure	Methotrexate exposure	Platinum exposure
Study	Wipe ID	Location	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)	exposure (ng/sq ft)	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)
Baseline	_	Hood tray	QN	>3578.6	2771.4	2,065.0	DN	DN	240.5
Baseline	2	Hood pass thru handle	QN	42.7	206.8	1,417.5	DN	DN	20.1
Baseline	ю	Negative pressure room interior door handle	Q	QN	77.0	66.6	QN	QN	QN
Baseline	4	Ante room interior door handle	QN	QN	QN	QN	QN	QN	QN
Baseline	5	Nurse's station counter	DN	QN	32.4	ND	DN	DN	DN
Baseline	6	Exterior ante room door handle	QN	QN	DN	ND	DN	DN	QN
Study I Wk I	_	Hood tray	365.9	56.9	2609.6	66.6	QN	DN	32.1
Study I Wk I	2	Hood pass thru handle	QN	Q	QN	ND	DN	DN	QN
Study I Wk I	e	Negative pressure room interior door handle	Q	QN	QN	QN	QN	QN	DN
Study I Wk I	4	Ante room interior door handle	QN	QN	QN	ND	QN	DN	QN
Study I Wk I	5	Nurses' station counter	QN	QN	QN	ND	QN	QN	QN
Study I Wk I	6	Exterior ante room door	DN	QN	QN	ŊD	DN	QN	QN
				!		!	!		
Study I Wk 2	_	Hood tray	67.2	QN	I 45.2	DN	DN	DN	30.9
Study I Wk 2	2	Hood pass thru handle	336.8	QN	DN	DN	DN	ND	DN
Study I Wk 2	m	Negative pressure room interior door handle	47.9	QN	60.6	DN	QN	QN	QN
Study I Wk 2	4	Ante room interior door handle	QN	QN	QN	ND	QN	DN	DN
Study I Wk 2	5	Nurses' station counter	QN	QN	QN	ND	DN	DN	QN
Study I Wk 2	9	Exterior ante room door	DN	QN	DN	QN	QN	DN	QN
Study 2	_	Hood tray	QN	Q	QN	ND	QN	DN	QN
Study 2	2	Hood airlock exterior handle	QN	DN	DN	ND	DN	DN	DN
Study 2	e	Negative pressure room interior door handle	Q	QN	QN	QN	QN	QN	DN
Study 2	4	Ante room interior door handle	QN	DN	DN	ND	DN	DN	DN
Study 2	5	Nurses' station counter	QN	Q	QN	DN	QN	DN	DN
Study 2	9	Exterior ante room door handle	QN	QN	DN	ND	DN	QN	DN
ND = non-detectable.	ble.								

			Docetaxel exposure	Paclitaxel exposure	5-FU Exposure	Cyclophos-phamide exposure	lfosfamide exposure	Methotrexate exposure	Platinum exposure
Study	Wipe ID	Location	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)	(ng/sq ft)
Baseline	_	Biological safety cabinet	QN	QN	59.48	22.2	QN	QN	QN
Baseline	2	Negative pressure room floor	DN	82.6	1460.9	108.1	QN	QN	46.4
Baseline	e	Ante room counter	DN	DN	47.4	DN	QN	QN	QN
Baseline	4	Negative pressure room interior door handle	QN	QN	Q	QN	Q	QN	QN
Baseline	5	Nurses' station counter	ND	DN	QN	DN	QN	QN	DN
Baseline	6	Exterior ante room door handle	QN	QN	QN	QN	QN	QN	QN
Study I Wk I	_	Biological safety cabinet	QN	QN	28.6	DN	QN	QN	QN
Study I Wk I	2	Negative pressure room floor	DN	DN	92.3	32.5	QN	QN	QN
Study I Wk I	e	Ante room counter	DN	QN	QN	QN	QN	QN	QN
Study I Wk I	4	Negative pressure room interior door handle	QN	QN	Q	QN	Q	QN	QN
Study I Wk I	5	Nurses' station counter	DN	DN	QN	QN	QN	QN	QN
Study I Wk I	6	Exterior ante room door handle	QN	QN	QN	ND	QN	QN	DN
Study I Wk 2	_	Biological safety cabinet	QN	QN	20.7	186.3	QN	QN	QN
Study I Wk 2	2	Negative pressure room floor	DN	DN	54.3	489.3	QN	QN	QN
Study I Wk 2	°	Ante room counter	DN	DN	QN	QN	QN	QN	QN
Study I Wk 2	4	Negative pressure room interior door handle	QN	DN	Q	QN	Q	Q	QN
Study I Wk 2	5	Nurses' station counter	DN	DN	QN	QN	QN	QN	QN
Study I Wk 2	6	Exterior ante room door handle	QN	QN	QN	ND	QN	DN	QN
Study 2	_	Biological safety cabinet	QN	QN	QN	DN	QN	QN	QN
Study 2	2	Negative pressure room floor	DN	DN	108.4	QN	QN	QN	QN
Study 2	e	Ante room counter	DN	DN	QN	QN	QN	QN	QN
Study 2	4	Negative pressure room interior door handle	QN	QN	Q	DN	Q	QN	QN
Study 2	5	Nurses' station counter	DN	DN	QN	QN	QN	QN	QN
Study 2	6	Exterior ante room door handle	DN	QN	QN	QN	QN	QN	QN

Table 9.	Table 9. Surface exposures of chemotherapeutic agents prior to and after the use of HDClean in the pharmacy and nursing unit at cancer center #3.	emotherapeutic	agents prior	to and after	the use of HD	Clean in the	pharmacy and	nursing unit	at cancer ce	nter #3.		
			Docetaxel exposure ng/ft² (ng/cm²)	:m²)	Paclitaxel exposure ng/ft ² (ng/cm ²)		5-FU exposure ng/ft ² (ng/cm ²)	e (Cyclophosphamide exposure ng/ft ² (ng/cm ²)	ohamide g/ft ²	lfosfamide exposure ng/ft ² (ng/cm ²)	g/ft ²
			Pre ^a	Post ^b	Pre ^a	Post ^b	Pre ^a	Post ^b	Pre ^a	Post ^b	Pre ^a	Post ^b
Wipe ID	Location description	Department	HDClean	HDClean	HDClean	HDClean	HDClean	HDClean	HDClean	HDClean	HDClean	HDClean
_	Pharmacy Hood 3 Middle	Pharmacy	15.7	Q	630.8 (0.68)	Q	135.1 (0.15)	QN	QN	QN	617.8	54.9
			(0.02)								(0.67)	(0.06)
7	Pharmacy Floor under Hood 3	Pharmacy	Q	QN	QN	QZ	Q	DN	QN	QN	Q	Q
e	Pharmacy Pass Through Window	Pharmacy	Q	QN	59.18 (0.06)	Q	QN	DN	DN	QN	Q	QN
4	Oncology Center – Pod 4 Counter	Pharmacy	Q	QN	QN	QZ	31.98 (0.03)	DN	DN	QN	QZ	QN
2	Oncology Center – Pod 4 Chemo Bin	Nursing	Q	QN	32.51 (0.04)	Q	QN	DN	DN	QN	Q	QN
6	Oncology Center – Pod 3 Counter	Infusion Area	Q	QN	QN	QN	11.28 (0.01)	QN	QN	QN	QN	Q
^a Pre HDCI. ^b Post HDC	^a Pre HDClean samples were obtained after the use of a CSTD and cleaning with Surface Safe. ^b Post HDClean samples were obtained after the use of CSTD and HDClean.	fter the use of a (after the use of C	CSTD and clea	aning with Surf Clean.	ace Safe.							

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areas were treated with Surface Safe prior to performing the wipe studies.

After the cleaning agent was changed to HDClean, all exposures of all drugs in all locations were nondetectable except for an exposure of ifosfamide in one location in the BSC (1 out of 30 = 3.3%) (P < 0.05). In addition, the exposure of ifosfamide at this location was reduced by 91.1% compared to baseline. The use of HDClean resulted in ~800% reduction in the number of locations with detectable drugs compared to the use of Surface Safe.

Additional studies were performed on areas with high use of cisplatin. The exposure of Pt at baseline prior to cleaning (n=4) and after cleaning with Surface Safe (n=4) were 479.5 ± 80.3 ng/sq ft and 503.5 ± 160.3 ng/sq ft, respectively. After cleaning with HDClean, the exposure of platinum (Pt) was non-detectable in all areas (n=4).

Discussion

This is the first series of research and development studies and testing in cancer centers that evaluated the ability of a cleaning product to remove surfaces contaminated with HDs with vastly different chemical characteristics and solubilities using analytical chemistry methods to accurately measure the concentrations of HDs on surfaces. HDClean was able to achieve pronounced reductions in all studies and complete decontamination in most studies that evaluated surface contamination of a wide range of HDs from a wide range of surfaces. In studies evaluating HDClean in two separate cancer centers that were not using a CSTD, daily use of HDClean was equal to or more effective in reducing surface contamination of HDs compared with a CSTD.¹⁹⁻²⁶ In addition, a third study in a separate cancer center was performed to evaluate the effectiveness of HDClean plus a CSTD compared with Surface Safe plus a CSTD. The results of this study demonstrated that HD residue remains on surfaces even when best practices are followed (e.g. use of CSTD). Moreover, the results of this study showed that HDClean is more effective in removing HDs from surfaces than Surface Safe and HDClean does not have the problems associated with Surface Safe (e.g. strong odor, corrosive to stainless steel).³⁰ In summary, the results of these studies fully support HDClean as an effective decontaminating and cleaning product for HDs based on the requirements outlined in the draft of USP800. Moreover, these are the first such studies published for any related product used to remove HD from surfaces.

HDClean was evaluated to remove the surface contamination of docetaxel, paclitaxel, 5-FU, ifosfamide, cyclophosphamide, and cisplatin that were added to 4 sq ft and 8 sq ft surface areas. The use of HDClean on the 4 sq ft area resulted in non-detectable concentrations of all drugs in all surface areas. The use of HDClean on the on the 8 sq ft area resulted in non-detectable concentrations of docetaxel, paclitaxel and cyclophosphamide in all surface areas and pronounced reductions in 5-FU and ifosfamide exposures. Thus, HDClean was equally effective in removing all drugs from the 4 and 8 sq ft areas with the exception of cisplatin where HDClean resulted in 100% and 60% reductions in the 4 sq ft and 8 sq ft areas, respectively. The results of this study show that HDClean is effective is decontaminating and cleaning a wide variety of HD from surfaces.

A separate study was also performed to evaluate the ability of HDClean to remove drug contaminations from various types of surfaces. In this study, daily of use HDClean after two days and especially five days resulted in pronounced reductions of all HD on all types of surfaces. In most cases, all of the drugs were non-detectable on all surfaces after use of HDClean for five consecutive days. This study showed that HDClean is effective in removing HDs from a wide range of surfaces that are commonly present in pharmacies and nursing units.

To evaluate the effectiveness of HDClean alone in actual pharmacies, it was tested under two different conditions in two cancer centers that did not use a CSTD. In study 1, the concentrations of most drugs at most locations after cleaning with HDClean were significantly reduced as compared to baseline even though the wipe studies were performed after the preparation of chemotherapy on that day but before the second HDClean procedure. In study 2, the surface exposure of all drugs on all surfaces were non-detectable except for 5-FU in 1 location in cancer center #2(detectable exposures in only 1.1% of locations) when HDClean was used to clean the surfaces at the end of preparing chemotherapy. These results of studies 1 and 2 highlight the need to use HDClean daily and especially at the end of the preparation of chemotherapy in order to remove all surface contamination associated with preparing chemotherapy that day. Moreover, the results of these studies suggest that HDClean is equal or more effective in reducing surface contamination of HDs compared with a CSTD.¹⁹⁻²⁸

A third study in a cancer center was performed to evaluate the effectiveness of HDClean plus a CSTD compared with Surface Safe plus a CSTD. These results of this study demonstrated that HD residue remains on surfaces even when all best practices are followed (e.g. use of CSTD). As per the prior studies, HDClean was able to fully remove drug exposures from surfaces contaminated with 5-FU, cyclophosphamide, docetaxel, and paclitaxel, and reduce the exposure of ifosfamide by >90%. Moreover, our results show that HDClean is more effective in removing a wide variety of HDs, especially platinum analogues, from various surfaces than Surface Safe.

Conclusion

HDClean is a multi-component system that fully addresses the guidelines outlined in USP800 for a product that can decontaminate and remove HDs from surfaces and overcomes the problems associated with sodium hypochlorite or bleach-like products. Based on this series of research and development studies, studies in cancer centers, and the broad range of chemical and solubility characteristics of the drugs tested, the results suggest that HDClean can be used to successfully remove a wide variety of HDs from surfaces in hospitals, pharmacies, nursing units and laboratories or any other place where HDs are present. Moreover, the results of our studies suggest that HDClean is more effective that Surface Safe in removing HDs, especially platinum analogues, and equal to or more effective than CSTD in controlling HD surface contamination.^{19-22,30} This study suggests that Pt analogues are much harder to remove from surfaces than other agents.

Potential limitations of this study include lack of evaluation of HDClean to remove surface exposures of biological agents (e.g. proteins and antibodies) and after acute chemotherapy spills. Additional studies are needed to address these types of exposures. However, the solutions used in HDClean are able to denature the types of proteins used in biological agents. For acute chemotherapy spills, it may be most appropriate to use existing methods to remove the chemotherapy and then use HDClean to clean the residual drug exposure. Additional studies are required to evaluate the factors associated with causing surface exposures, removing surface exposures, especially Pt agents, use of HDClean over larger surface areas, and to document potential safe levels of exposure.

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